



# Polymer Dynamics: From Synthetic Polymers to Proteins

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# Collaborators







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B. Brockhouse Chalk River Canada

Nobel prize 1994



C. Shull Oak Ridge USA Neutrons tell where the atoms are and how they move

**Neutrons and Polymers** 





Centre for Neutron Science

Neutrons access molecular length and time scales

Neutrons see the Nuclei





### Only neutrons enable isotropic contrasting

# Polymer Chain Conformation



Early triumph of neutron scattering





# Polymer

Dynamics





#### **Pair Correlations**



**Collective Dynamics** 

# Self Correlation



#### Self motion

# Observation of polymer motion





# Standard Model

# Rouse Dynamics



Gaussian chain





Chain in a heat bath
 Thermally activated fluctuations
 Relaxation by viscous and entropic forces
 No explicit chain-chain interactions



#### Gaussian chain entropic force

#### mean square segment displacement

$$S(Q,t) = \exp\left[-\frac{Q^2}{6}\langle r^2(t)\rangle\right]$$

$$\left\langle \boldsymbol{r}^{2}\left(\boldsymbol{t}\right)\right\rangle \approx\boldsymbol{t}^{0.5}$$

#### **Neutron Spin Echo**



### Sublinear diffusion of chain segments



# Segment Dynamics in the Melt



Rouse prediction verified





# Reptation







P.G. De Gennes ESPCI Paris France

#### Nobel prize 1991

### Subject of intense current research





**Coherent scattering: form factor of the tube** 

# Topological Constraint Motion - Self-Motion



**Centre for Neutron Science** 

Mean squared displacement





All parameters from single chain structure factor





# Reptation

# Limiting





Limiting Processes

### Contour length fluctuation

#### **Constraint release**



### **Reptation inherent mechanisms**



# Separation of Contour Length Fluctuations (CLF)



![](_page_19_Picture_3.jpeg)

$$\psi(t) = C_{\mu} \frac{N_{e}}{N} \left(\frac{t}{T_{e}}\right)^{\frac{1}{4}}$$

number of segments released from tube

A.E. Likhtman et al., Macromolecules (2002)

![](_page_20_Figure_0.jpeg)

Inner part moves as infinite chain

![](_page_21_Picture_0.jpeg)

**Constraint Release** 

![](_page_21_Picture_2.jpeg)

Common approach CR relates to chain diffusion only

Here: CLF may play important role

Zamponi et al. PRL 2006

### Chains move away and open tube laterally

![](_page_22_Picture_0.jpeg)

**Constraint Release** 

![](_page_22_Figure_2.jpeg)

#### Loss of confinement with decreasing matrix length

![](_page_23_Picture_0.jpeg)

# What are the Processes behind CR?

![](_page_23_Picture_2.jpeg)

![](_page_23_Picture_3.jpeg)

CR needs a confining chain to diffuse away; time scale:

reptation time of matrix chain

e.g. 12K τ<sub>d</sub> = 5000 ns

We observe strong effects on a scale of 200ns!

![](_page_24_Picture_0.jpeg)

# Experiment to Separate Processes

 a) Long chain in short matrix no CLF of long chains
 Constraint release only (end part is small)

![](_page_24_Picture_3.jpeg)

b) Short chain in long matrix no CR by long chains 12/36
Contour length fluctation only (long chains do not diffuse on our time scale)

![](_page_25_Figure_0.jpeg)

![](_page_25_Figure_1.jpeg)

![](_page_25_Figure_2.jpeg)

#### long in short

CR through not by diffusion (5000ns) but by CLF of short chains

![](_page_25_Figure_5.jpeg)

# CR may be caused solely by CLF

![](_page_26_Picture_0.jpeg)

Topological Constraints: Summary

![](_page_26_Figure_2.jpeg)

### NSE and synthetic chemistry

- Direct visualization of
  Regime of local reptation
  Limiting mechanisms
  - contour length fluctuations
  - constraint release

Basic processes for the hierarchical relaxation of branched polymers

![](_page_26_Picture_8.jpeg)

![](_page_27_Picture_0.jpeg)

![](_page_27_Picture_1.jpeg)

# Dynamics of **Biopolymers:** Large Scale Motion

![](_page_28_Picture_0.jpeg)

![](_page_28_Figure_1.jpeg)

Protein motion and the coordination of biofunction

- Genome regulatory proteins
- Motor proteins
- Signaling proteins
- Structural proteins

![](_page_28_Picture_7.jpeg)

#### Are large scale thermal fluctuations related to functional dynamics?

![](_page_29_Picture_0.jpeg)

Alcohol Dehydrogenase

Important enzyme e.g. prohibits poisoning by Italian red wine

In humans (Dimers) Catalysis oxidation of ethanol

 $\label{eq:ch_3} \begin{array}{l} \mbox{CH}_2 \mbox{ OH } + \mbox{ NAD}^{\scriptscriptstyle +} \rightarrow \mbox{CH}_3 \mbox{ CHO } + \\ \mbox{NADH } + \mbox{ H}^{\scriptscriptstyle +} \end{array}$ 

![](_page_29_Picture_5.jpeg)

![](_page_30_Picture_0.jpeg)

# Alcohol Dehydrogenase

#### In yeast (Tetramer) Fermentation process

 $\begin{array}{l} \mathsf{CH}_3 \ \mathsf{CHO} \ + \ \mathsf{NADH} \ + \ \mathsf{H}^+ \ \rightarrow \ \mathsf{CH}_3 \ \mathsf{CH}_2 \\ \mathsf{OH} \ + \ \mathsf{NAD}^+ \end{array}$ 

![](_page_30_Figure_5.jpeg)

![](_page_30_Picture_6.jpeg)

#### **Reverse** action

![](_page_31_Picture_0.jpeg)

Alcohol Dehydrogenase

![](_page_31_Figure_2.jpeg)

#### **Dynamics**

- Internal aggregate motion
- Rotational dynamics
- Influence of cofactor NADH

![](_page_31_Figure_7.jpeg)

![](_page_31_Figure_8.jpeg)

![](_page_32_Picture_0.jpeg)

![](_page_32_Figure_1.jpeg)

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#### structure factor

![](_page_32_Figure_3.jpeg)

#### concentration scaled

![](_page_33_Picture_0.jpeg)

Solution Structure

![](_page_33_Figure_2.jpeg)

**DAMMIN** program

![](_page_33_Figure_4.jpeg)

Solution structure differs from crystal: crossed dimers

![](_page_34_Picture_0.jpeg)

![](_page_34_Picture_1.jpeg)

![](_page_34_Picture_2.jpeg)

![](_page_35_Picture_0.jpeg)

![](_page_35_Figure_2.jpeg)

Dynamic light scattering: photon correlation

Neutron Spin Echo Results

![](_page_36_Figure_1.jpeg)

ADH + NAD at 5% concentration

![](_page_37_Picture_0.jpeg)

# Effective Diffusion Coefficient

![](_page_37_Figure_2.jpeg)

![](_page_37_Figure_3.jpeg)

- Strong Q modulation
- Significant conc.
   dependence
- At low Q agreement with light scattering

Important contributions beyond trans. diffusion

![](_page_38_Picture_0.jpeg)

# Low Q Data: Interaction

![](_page_38_Figure_2.jpeg)

 $D_{eff} = D_0 \frac{\pi}{S}$ 

![](_page_39_Picture_0.jpeg)

Low Q Data: Interaction

![](_page_39_Figure_2.jpeg)

$$\boldsymbol{D}_{eff} = \boldsymbol{D}_{0} \frac{\boldsymbol{H}(\boldsymbol{Q})}{\boldsymbol{S}(\boldsymbol{Q})}$$

![](_page_39_Figure_4.jpeg)

![](_page_40_Picture_0.jpeg)

Corrected D<sub>eff</sub>(Q) by S(Q)

# S(Q) correction removes low Q structure

0,7

0,5

1.5

 $Q/nm^{-1}$ 

2

![](_page_41_Picture_0.jpeg)

# Low Q Data: Interaction

![](_page_41_Picture_2.jpeg)

![](_page_41_Figure_3.jpeg)

$$D_{eff} = D_0 \frac{H(Q)}{S(Q)}$$

#### H(Q) not known

First approximation Both H(Q) and S(Q) by Percus Yevic appr. of hard spheres

![](_page_42_Picture_0.jpeg)

# Low Q Data: Interaction

![](_page_42_Picture_2.jpeg)

![](_page_42_Figure_3.jpeg)

![](_page_42_Figure_4.jpeg)

Qualitatively correct

Low Q: very little conc. dependence

**Entermediate Q:** reduction slightly weaker than exp.

Higher Q: no conc. dependence

![](_page_43_Picture_0.jpeg)

![](_page_43_Picture_1.jpeg)

First cumulant approach:  $Q^2 D_{eff} = \overline{\Gamma}(Q) = -\lim_{t \to 0} \frac{\partial}{\partial t} \frac{S(Q,t)}{S(Q,0)}$ 

$$D_{eff}(Q) = \frac{k_B T}{Q^2} \frac{\sum_{j,k} \left\langle b_j e^{iqr_j} \begin{pmatrix} Q \\ Q \times r_j \end{pmatrix} \widehat{H} \begin{pmatrix} Q \\ Q \times r_k \end{pmatrix} b_k e^{-iqr_k} \right\rangle}{\sum_{j,k} \left\langle b_j e^{iqr_j} \ b_k e^{-iqr_k} \right\rangle}$$

<u>r</u><sub>j</sub>, <u>r</u><sub>k</sub> atom positions; b<sub>j</sub> b<sub>k</sub>: scattering length denominator: structure factor

![](_page_44_Picture_0.jpeg)

![](_page_44_Picture_1.jpeg)

First cumulant approach:  $Q^2 D_{eff} = \overline{\Gamma}(Q) = -\lim_{t \to 0} \frac{\partial}{\partial t} \frac{S(Q,t)}{S(Q,0)}$ 

#### Programme HYDROPRO: calculates the full mobility matrix H

![](_page_44_Figure_4.jpeg)

Garcia de la Torre, Biophysical J. <u>78</u>, 719 (2000)

![](_page_45_Picture_0.jpeg)

#### Very sensitive to correct solution structure!!

![](_page_45_Figure_2.jpeg)

translational + rotational diffusion

### HYDROPRO result scaled to low Q trans. diffusion

![](_page_46_Picture_0.jpeg)

# High Q Data: Internal Motions

![](_page_46_Figure_2.jpeg)

![](_page_46_Picture_3.jpeg)

# Coupling term

- elongational coupling
- Rotational coupling of domain motion

![](_page_46_Figure_7.jpeg)

![](_page_47_Picture_0.jpeg)

# High Q Data: Internal Motions

![](_page_47_Figure_2.jpeg)

- Very consistent data sets at different concentration and cofactor
- Quantitative description by rotational diffusion of crossed dimers and rotational coupling
- NAD leads to slowing down in the low Q-flank
  - significant effect
  - points to collective origin

### All data scaled with translational diffusion

![](_page_48_Picture_0.jpeg)

Alcoholdehydrogenase Dynamics

![](_page_48_Figure_2.jpeg)

Structure in solvent different from crystal structure

Rigid body dynamics dominates but clear evidence for internal motion

### Influence of cofactor

![](_page_49_Picture_0.jpeg)

![](_page_49_Picture_1.jpeg)

Solvent Structure

**Influence** of Cofactor

**Aggregate Motion**